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Pathologies associated to the consumption of aflatoxin M₁ in dairy products Patologías asociadas al consumo de aflatoxina M₁ en productos lácteos

Araceli Monter-Arciniega^a, Nelly del Socorro Cruz-Cansino^b, Esther Ramírez-Moreno^c, Quinatzin Y. Zafra-Rojas^d, Alicia Cervantes-Elizarrarás^e, José Alberto Ariza-Ortega^f

Abstract:

Milk is the most traded, processed and consumed dairy product in the world, approximately 113.7 to 150 kg per capita. However, one of the food safety and human health problems worldwide is aflatoxin M_1 (AFM₁), present exclusively in milk and its derivatives, because this mycotoxin is heat-resistant to common food industrialization processes and is capable of producing hepatotoxicity, carcinogenicity, genotoxicity and immunosuppression in humans, so there are international regulations that establish parameters ranging from 0.03 to 0.250 µg/kg depending on each country. The present review aims at compiling information on mycotoxins and specifically on AFM₁ from its biotransformation from dairy cattle until the pathologies in humans associated with its consumption.

Keywords:

Aflatoxin M1, Milk, Pathologies

Resumen:

La leche es el producto lácteo más comercializado, elaborado y consumido en el mundo, aproximadamente de 113.7 a 150 kg *per cápita*. Sin embargo, uno de los problemas de seguridad alimentaria y salud humana a nivel mundial es la aflatoxina M_1 (AFM₁), presente exclusivamente en la leche y sus derivados, debido a que esta micotoxina es termorresistente a procesos comunes de industrialización de alimentos y es capaz de producir hepatotoxicidad, carcinogenicidad, genotoxicidad e inmunosupresión en humanos, por lo que existen regulaciones internacionales en donde establecen parámetros que van de 0.03 a 0.250 µg/kg dependiendo de cada país. La presente revisión tiene como objetivo recopilar información sobre las micotoxinas y específicamente sobre la AFM₁ desde su biotransformación a partir del ganado lechero, hasta las patologías en el ser humano asociadas a su consumo.

Palabras Clave:

Aflatoxina M1, Leche, Patologías

^a Área Académica de Nutrición, Instituto de Ciencias de la Salud, Universidad Autónoma del Estado de Hidalgo, San Agustín Tlaxiaca, Hidalgo, México. https://orcid.org/0000-0003-1475-8093, Email: mo270469@uaeh.edu.mx

- ^b Corresponding author, Área Académica de Nutrición, Instituto de Ciencias de la Salud, Universidad Autónoma del Estado de Hidalgo, San Agustín Tlaxiaca, Hidalgo, México. https://orcid.org/0000-0002-6771-3684, Email: ncruz@uaeh.edu.mx
- ^c Área Académica de Nutrición, Instituto de Ciencias de la Salud, Universidad Autónoma del Estado de Hidalgo, San Agustín Tlaxiaca, Hidalgo, México. https://orcid.org/0000-0002-9928-8600, Email: esther_ramirez@uaeh.edu.mx.
- ^d Área Académica de Nutrición, Instituto de Ciencias de la Salud, Universidad Autónoma del Estado de Hidalgo, San Agustín Tlaxiaca, Hidalgo, México. https://orcid.org/0000-0002-5295-9972. Email: quinatzin_zafra@uaeh.edu.mx.
- ^e Área Académica de Nutrición, Instituto de Ciencias de la Salud, Universidad Autónoma del Estado de Hidalgo, San Agustín Tlaxiaca, Hidalgo, México. https://orcid.org/0000-0002-1432-2882, Email: <u>alicia_cervantes@uaeh.edu.mx</u>.
- ^f Área Académica de Nutrición, Instituto de Ciencias de la Salud, Universidad Autónoma del Estado de Hidalgo, San Agustín Tlaxiaca, Hidalgo, México. https://orcid.org/0000-0002-2163-4593, Email: jose_ariza@uaeh.edu.mx.



INTRODUCTION

Aflatoxins (AF) are toxic compounds produced as secondary metabolites of different toxigenic fungi such as Aspergillus flavus and Aspergillus parasiticus, which produce AFB₁ in feed and food crops,1 when these are consumed by dairy cattle or some other lactating animal, the digestive system converts part of this AF into AFM₁, with a conversion rate of 0.3 to 6.2%,² to be subsequently excreted in milk (approximately 0.05 µg/kg, at an intake <40 µg per day per dairy animal fed contaminated feed), because the consumption of this feed is important to the majority of the population, it may pose a risk to human health,³ since although AFM1 is less carcinogenic (10% in vivo) than AFB₁.⁴ In addition, pathologies such as hepatotoxicity, carcinogenicity, cytotoxicity, genotoxicity, and immunosuppression have been reported due to its consumption, making it of special concern in terms of food safety and human health.5 Outbreaks of mycotoxicosis due to aflatoxins consumption, cause hundreds of deaths that can be prevented or interrupted by food analysis and treatment.⁶ AF are regulated in more than 80 countries, but their legislation is not known at the international level; therefore, only permitted limits have been established for AFM1 in milk and dairy products, ranging from 0 to 1.0 g/kg.⁷ The objective of this review article is to summarize information such as definition, biotransformation, reports of aflatoxin M₁ presence in milk and pathologies associated with its consumption.

MYCOTOXINS

Mycotoxins are toxic compounds, classified as chemical hazards of biological origin, which are produced naturally by some species of molds belonging to the genera Aspergillus, Penicillium, Fusarium, Claviceps and, Alternaria; these mycotoxins are usually formed at the end of the exponential phase or the beginning of the stationary phase of mold growth, during storage or in the food itself. When found in warm and humid places, before or after harvest, contaminate food, feed, or raw materials used in their processing. These compounds causes diseases and disorders called mycotoxicosis, which has been shown to cause damage to the health of both humans and animals, these effects are caused by inhalation, direct contact, or ingestion. The most frequent mycotoxins that pose a risk are AF, achroatoxin A, patulin, fumonisins, zearalenone, nivalenol, and deoxynivalenol.8,9 The first recorded data on mycotoxicosis date back to the Middle Ages and correspond to the disease called Ergotism, better known as St. Anthony's Fire, which affected a large number of people in Europe and was caused by the consumption of rye contaminated with alkaloids produced by *Claviceps purpurea*.¹⁰ On the other hand, *Aspergillus flavus*, has its first record in the United Kingdom in 1960, where the death of more than 100 thousand turkeys fed with feed based on peanut flour, which came from Brazil and which contained the secondary metabolites of the fungal species,11 these metabolites were later denominated as AF and are the first compounds identified as such.⁸

AFLATOXINS

AF are classified as non-protein organic compounds, are secondary metabolites produced by Aspergillus flavus and Aspergillus parasiticus fungi, which require a minimum temperature of 6 to 8 °C, optimum of 36 to 38 °C and maximum of 44 to 46 °C for their growth, these fungal strains produce AF in conditions of 12 °C as minimum temperature, ²⁷ to 30 °C as optimum temperature and 40 to 42 °C as maximum ¹², pH of 5, water activity less of 0.70 at a relative humidity of 80%, and A. flavus produces the highest amount of toxins.13 AF are characterized by having the form of solid crystals ranging in color from white to yellow. Chemically are isocoumarin derivatives, whose basic skeleton is a furan ring attached to the coumarin nucleus (Figure 1), the molecular weight is between 312.06 and 346.06 g/mol approximately, have low solubility in water (10-30 µg/ml), but in organic solvents such as chloroform, ethanol, methanol, acetonitrile, and acetone are soluble, also not very stable to light and air when in their pure form, tend to be susceptible to alkaline hydrolysis and contact with ammonia or sodium hypochlorite solutions (pH >10.5), and their ability to adapt to heat makes them heat-resistant and at pH 3 and 10 maintain their stability.¹⁴⁻¹⁷ AF is produced by A. flavus and A. parasiticus molds growing in soil, decaying vegetation and cereals, which are among the most toxic mycotoxins, mostly related to A. flavus. The most affected crops are cereals, oilseeds, and spices.^{9,18} AF belong to a wide group of approximately 20 different types, however, the most common are B_1 , B_2 , G_1 and G_2 , according to the blue or green fluorescence in the presence of ultraviolet light at 365 nm, AFM₁ has also been included as one of the mycotoxins of importance in human health.¹⁵ The association of AF ingestion and the incidence of pathologies, mainly hepatocellular carcinoma (HCC).¹⁹ One of the first cases presented, pointing to the association between the incidence of hepatocellular carcinoma (HCC) with the ingestion of AF.¹⁹, was presented in studies conducted in China and sub-Saharan Africa,^{20,21} which formed the basis for AF to be classified as a human carcinogen by the International Agency for Research on Cancer (IARC) since 1994.22

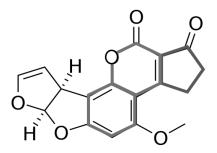


Figure 1. Basic structure of aflatoxins. Furan ring. Coumarin ring.

TOXICITY OF AFLATOXINS

The toxicity of AF in humans is determined by different factors such as the bioavailability and toxicity of the mycotoxin, the synergisms between them, the amount of mycotoxin ingested daily depending on its concentration, the continuity or intermittent consumption of the contaminated food, the weight, physiological state, health and age of the individual.⁹ The toxigenic effect of AF generate alteration in the synthesis of DNA, RNA, and proteins, which causes the appearance of changes in the permeability of the mitochondrial membrane, alteration of ribosomes, decrease in cellular respiration, and interruption of electron transport,²³ toxicity is the result of generalized and unspecific interactions between AF and diverse cellular proteins, which can cause the interruption of basic metabolic processes, protein synthesis and cause cell death.²⁴ The dose and duration of exposure to AF has been shown to play an important role in toxicology, exposure to large doses (acute aflatoxicosis) can cause pathologies and even death, while high-level exposure causes acute hepatic necrosis, which develops sequelae of cirrhosis or HCC.25 As mentioned above, one of the mycotoxins of importance is AFM₁, which reaches milk through the metabolization of AFB1 in dairy cattle consuming AFB₁ contaminated food or feed.

BIOTRANSFORMATION OF AFLATOXIN M1

After ingestion by dairy cows through contaminated feed, AFB₁ is partially metabolized and bio-transformed to AFM₁.²⁶ A part of AFB₁ consumption by dairy cattle is transformed by some microorganisms to form aflatoxicol (highly toxic).27 The remaining part that is not degraded by the rumen is absorbed in the small intestine because it has lipophilic properties and is of low molecular weight and then transported through the portal bloodstream.²⁸ Once it reaches the liver, it is metabolized by mixed-function microsomal oxidase enzymes (CYP450 superfamily enzyme) to a reactive epoxide intermediate (8,9epoxide), which is responsible for DNA mutation and can bind RNA and proteins, which can lead to cellular dysregulation.²⁵ This hepatic biotransformation is also responsible for subjecting AFB₁ to reduction, epoxidation, hydroxylation, and demethylation. The hydroxylation phase produces AFM₁,^{29,30} which is subsequently distributed to the mammary gland via the bloodstream, to be excreted in the milk.31,32 This excretion may take place by passive diffusion, but active transport, mediated by outward transporters of the ABC family, expressed in the epithelial cells of the mammary gland, is probably the principal means of excretion.33

CHARACTERISTICS OF AFLATOXIN M1

AFM₁, the 4-hydroxy-metabolite of fungal AFB₁, is excreted in the milk of all lactating species, including cows.³⁴ This AF has a chemical structure similar to AFB₁, but with an OH group attached to the tetrahydrofurans (Figure 2) and possesses toxic effects even at low concentrations (≤ 1 ng/kg/day).³⁴ Its molecular weight is 328 g/mol and it has a melting point of 299 °C is resistant to temperatures between 260 and 320 °C without decomposing,³⁵ and remains stable in milk and dairy products after industrial processes, such as pasteurization and ultrapasteurization. It has been mentioned that AFM₁ may be associated with the casein fraction of milk, because it increases its hydrophobicity during the proteolysis stage in cheese making, this condition is necessary for AFM₁ to bind.^{4,36} It was previously mentioned that according to the IARC, mycotoxins are found in group 1, the same agency classifies AFM₁ as a possible human carcinogen, which indicates that there is some evidence of causing cancer in humans but it has not yet been conclusive,37 therefore, different countries have established maximum permissible limits for AFM₁ in milk and milk products (Table 1), in order to carry out pertinent regulations, since the evidence indicates the presence of this mycotoxin in products in order to carry out regulations, since the evidence points to the presence of this mycotoxin in products of popular consumption such as liquid milk (Table 2). The range of contamination goes from 0. 009-1020.00 ng/L, 0.01-5.16 µg/kg, 9.0-12.00 pg/kg and 0.53-0.207 ppb. Mexico has an established limit of 0.05 µg/kg and according to the AFM₁ content in raw milk reported by Rangel-Muñoz et al.,38 their results exceed what is established in the country. Considering the maximum permissible limit of the European Union (0.05 µg/kg), countries such as Africa, Brazil, China, Egypt, El Salvador, Spain, Ethiopia, India, Iran, Malawi, Mexico, Pakistan, and Serbia exceed this regulation. In addition to the fact that they are close to the TD50 (10.38 µg/kg body weight/day),³⁹ the substantial public health problem and the possible development of pathologies caused by AFM₁ consumption are exposed.

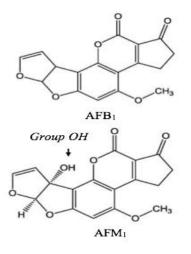


Figure 2. Difference between AFB₁ and AFM₁ in their chemical structure.

Table 1. Maximum permissible limit of AFM1 in different countries.

Country	Maximum limit µg/kg	Food
Austria	0.05 ^a	Milk and dairy products
Belgium	0.05 ^a	Milk
Bulgaria	0.50ª	Milk
China	15.00ª	Milk and dairy products
Czech Republic	0.10^{a} 0.50^{a}	Milk for children Milk for adults
European Union	0.05^{b} 0.025^{b}	Milk Infant and follow-on formula
France	0.03^{a} 0.05^{a}	Milk for children Milk for adults
Germany	0.05ª	Milk
Honduras	0.02^{a} 0.05^{a} 0.25^{a}	Baby food Dairy products Cheese
Hungary	0.05 ^a	Milk and dairy products
India	30.00 ^a	Milk and dairy products
Indonesia	5.00 ^a	Milk cheese
Iran	0.05^{a} 0.50^{a} 0.01^{a} 0.20^{a}	Pasteurized, raw or sterilized milk and milk derivatives Powdered milk Baby milk Cheese
Israel	0.05 ^a	Milk and dairy products
Italy	0.01 ^a	Baby food
Korea	0.05 ^a	Milk and dairy products
Latvia	0.50^{a}	Milk and dairy products
Moldova	0.50 ^a	Milk
Malta	0.05ª	Milk
Mercosur	0.50ª 5.00ª	Liquid milk Powdered milk

Mexico	0.05ª	Milk, milk formula and combination products
Morocco	0.05^{a} 0.03^{a} 0.50^{a} 0.30^{a}	Dairy products Milk for < 3 years Powdered milk Powdered milk for < 3 years
Netherlands	0.05^{a} 0.02^{a} 0.20^{a}	Milk Butter Cheeses
Peru	0.50 ^a	Milk
Poland	0.05ª	Dairy products
Romania	0.50ª	Milk and dairy products
Russia	0.50 ^a	Milk and dairy products
Saudi Arabia	0.20 ^a	Milk and dairy products
Senegal	0.50 ^a	Milk and dairy products
Singapore	0.50ª	Dairy products
Slovakia	0.50^{a} 0.10^{a}	Milk and dairy products Infant formulas and milk-based baby foods
Switzerland	0.05^{a} 0.25^{a} 0.02^{a}	Milk and dairy products Cheese Infant and follow-on formula
Sweden	0.050ª	Baby food
Syrian Arab Republic	0.20^{a} 0.05^{a}	Liquid milk Powdered milk
Taiwan Province of China	0.50^{a} 5.00^{a}	Liquid milk Powdered milk
Turkey	0.05^{a} 0.5^{a} 0.25^{a} 0.05^{a}	Liquid milk Powdered milk Cheese Milk-based baby food
Ukraine	0.50 ^a	Milk, dairy products y baby food

United States	0.50ª	Milk
Venezuela	0.50 ^a 5.00 ^a	Liquid milk Powdered milk
Vietnam	0.50ª	Milk and dairy products

^a: FAO Food and Nutrition Paper,⁴⁰. ^b: Commission Regulation (EC),⁴¹

Table 2. Presence of AFM_1 in milk in different countries.

Table 2. Presence of AFM ₁ in milk in alfferent countries.			
Country	Quantity	Food	
Africa	0.05–0.1 µg/kg. ⁴²	Milk	
Africa	4.8–26.11 ng/kg.43	Milk	
Algeria	95.59–557.22 ng/L. ⁴⁴	Raw milk	
Bangladesh	22.79–1489.28 ng/kg 18.11–672.18 ng/kg 25.07–48.95 ng/kg. ⁴⁵	Raw milk Pasteurized milk UHT milk	
Brazil	0.01–0.81 μg/kg. ⁴⁶	Commercial milk	
Brazil	12.84–18.72 ng/L. ⁴⁷	Raw milk	
Brazil	150–1020 ng/kg. ⁴⁸	UHT milk	
Chile	0.015 µg/kg. ⁴⁹	Milk	
China	$0.022 – 0.049 \ \mu g/kg^{50}$	Milk	
China	25.5-60.0 ng/L. ⁵¹	Raw milk	
Costa Rica	0.042-154 ng/mL. ⁵²	Raw milk	
Ecuador	0.023–0.751 µg/kg. ⁵³	Raw milk	
Egypt	0.02–0.19 µg/kg. ⁵⁴	Raw milk	
Egypt	0.053–0.207 ppb. ⁵⁵	Raw milk	
Egypt	0.18–0.41 µg/L. ⁵⁶	Raw milk	
Egypt and South Africa	8.52–78.06 ng/L. ⁵⁷	Commercial milk	
El Salvador	22.5–10.3 µg/kg. ⁵⁸	Raw milk	
Ethiopia	0.031–5.16 µg/L. ⁵⁹	Raw milk	
India	0.013–0.396 μg/L. ⁶⁰	Milk	
India	0.027–2.281 μg/kg 0.01–1.116 μg/kg. ⁶¹	Pasteurized milk	

		Commercial milk
India	0.598–1.29 µg/L. ⁶²	Raw milk
India	0.80–3.2 μg/L. ⁶³	Commercial milk
Iran	0.005–0.098 μg/kg 0.005–0.120 μg/kg. ⁶⁴	UHT milk Pasteurized milk
Iran	0.011–0.321 µg/L. ⁶⁵	Raw milk
Iran	10–150 ng/L. ⁶⁶	Commercial milk Raw milk
Iran	11.7–106.6 ng/L. ⁶⁷	Pasteurized milk
Italy	0.009–0.015 μg/kg. ⁶⁸	Raw milk
Italy	0.009–0.026 ng/Kg. ⁶⁹	Commercial milk
Italy	0.71–3.63 ng/kg 0.85–44.39 ng/Kg. ⁷⁰	UHT Milk Pasteurized milk
Italy	7.19–22.53 ng/kg. ⁷¹	Raw milk
Kenya	290.3–500 ng/kg. ⁷²	Commercial milk
Malawi	$0.42 - 1.02 \ \mu g/L.^{73}$	Raw milk
Mexico	0.021–18.5 µg/kg. ³⁸	Raw milk
Mexico	9.0–12.00 pg/mL. ⁷⁴	Raw and pasteurized milk
Pakistan	0.03 ng/ml 0.35 ng/ml 0.11 ng/ml. ⁷⁵	Raw milk UHT milk Pasteurized milk
Pakistan	0.187–0.346 μg/L. ⁷⁶	Raw milk Processed milk
Pakistan	0.3–1.0 µg/L. ⁷⁷	Raw milk
Serbia	0.012–0.53 μg/L. ⁷⁸	Raw milk
Serbia	0.024–0.319 μg/kg 0.038–0.231 μg/kg. ⁷⁹	UTH milk Pasteurized milk
Serbia	0.076–0.069 µg/kg. ⁸⁰	Raw milk

Spain	$0.05{-}0.50~\mu\text{g/L}.^{81}$	Commercial milk
Spain	9.0–18.8 ng/kg. ⁸²	Raw milk
Uruguay	0.005–0.08 µg/L. ⁸³	Commercial milk

PATHOLOGIES CAUSED BY AFLATOXIN M1

AFM₁ can cause DNA damage, genetic mutation, chromosomal abnormalities, and cell transformation in mammals *in vitro*, in insects, lower eukaryotes, and bacteria.³ The metabolic and toxic effects are mainly observed in the liver,^{84,85} although it has also been reported in the lung after exposure by inhalation and diet, and it is also a pathogenic factor in weight insufficiency, hypo-immunity, neurological damage, and even high infant mortality,^{86,87} in general, the main repercussions of AFM₁ are hepatotoxicity, carcinogenicity, genotoxicity, and immunosuppression.

Hepatotoxicity. Although AFM₁ is less mutagenic and genotoxic than AFB₁, it possesses cytotoxicity in hepatocytes *in vitro* and it has been demonstrated that its acute toxicity in several species is similar to that of AFB₁,³ in addition this AF also possesses hepatotoxic effects and is relatively stable in the processes of pasteurization, storage and processing of milk.⁸⁸ Marchese et al.,⁸⁹ highlights damage in HepG2 cells (human hepatoma cell line), carrying out the analysis of the effects of AFM₁ on the metabolomic and cytochemical profile in a hepatoblastoma cell line, it was found that AFM₁ is able to block the cell cycle in the G0/G1 phase, it also induces the modulation of lipid, glycolytic and amino acid metabolism, increases the levels of proinflammatory cytokines such as IL-6, IL-8 and TNF- α and decreases the levels of IL-4.

Carcinogenicity. Although AF ingestion has classically been associated with primary liver cancer or HCC and bile duct hyperplasia,⁹⁰ organs such as kidney, pancreas, bladder, bones, viscera, among others, have also been reported to develop cancer after mycotoxin exposure.91 The carcinogenic effect of AFM₁ metabolites is produced from DNA intercalation and base alkylation by the epoxide moiety, resulting in mutations in the p53 gene, which is important in preventing cell cycle progression in DNA mutations or programmed cell death signaling.²⁵ The risk of cancer due to exposure to the various forms of AF is based on cumulative lifetime dose, thus IARC identifies AFM1 as carcinogenic (group 2B), therefore, its regulation is limited to very low concentrations, this mycotoxin can cause cancer after prolonged exposure or at high doses (≥1 ng/kg/day), such as liver cancer and breast cancer (causing cell death, cell cycle detection and apoptosis), which are the most

common causes of death in men and women, respectively.⁹²⁻⁹⁴ On the other hand, taking into account the findings of Creppy, ⁹⁵ in the weighted average of consumption, only an intake of <1 ng/body weight contributes to the risk of liver cancer, as indicated by the Joint FAO/WHO Expert Committee on Food Additives (JECFA)⁹⁶ so it could be said that the development of pathologies associated with AFM₁ would be present almost everywhere in the world.

Cytotoxicity. A study in vitro reported that AFM₁ causes toxicity in human MCL-5 lymphoblastoid B cells and cHol cells, expressing or not human cytochrome P450 enzymes.⁹⁷ One study found that AFM₁ did not significantly inhibit the differentiated or undifferentiated phase of Caco-2 cell growth when membrane damage was detected simultaneously by monitoring lactose dehydrogenase (LDH) release.⁹⁸ In models of differentiated and undifferentiated Caco-2 cells, where they were exposed to a dose range of 0. 01 to 1 mg/mL of AFM₁, caused marked cytotoxicity and decreased survival, furthermore, this mycotoxin also caused cell membrane damage,^{99,100} indicating that it could damage intestinal cells at the cell membrane and affect DNA integrity, evidencing that, prolonged consumption of AFM1 containing foods may pose a risk of injury to the intestinal tract,⁹⁹ as reported by Gao et al.,¹⁰¹ on the toxicity of AFM₁, on the intestinal barrier, resulting in dysfunction of intestinal integrity in Caco-2 cells differentiated through clathrin-mediated endocytosis into TJ proteins. Huiying et al.,¹⁰² investigated the effects of AFM₁ on renal cells, showed that long-term administration of AF, causes renal damage through oxidative stress and apoptosis. On the other hand, it was analyzed whether L-proline alleviates renal injury caused by AFM₁ in a mouse model, to which different mycotoxins were administered, including AFM1 at dose of 3.5 mg/kg, the mycotoxin in question, activates pathways related to oxidative stress and causes renal damage; however, L-proline significantly alleviated oxidative damage and decreased apoptosis.¹⁰³ Nan et al.,¹⁰⁴ studied lactoferrin as an inhibitor of AFM₁-induced cytotoxicity and DNA damage in Caco-2, HEK, HepG2 and SK-N-SH cells, AF inhibited cell growth, induced an increase in lactate dehydrogenase level, as well as caused genetic damage and evidenced cytotoxicity and DNA damage, in the mentioned cell lines. Yanan et al.,¹⁰⁵ with Caco-2 cells, analyzed the expression of inflammation-related genes, revealed the inflammatory response caused by mycotoxins such as AFM₁, and demonstrated a decrease in gene expression of proinflammatory cytokines. Finally, the toxicity of AFM₁ in human cell lines has been shown to not require CYP450 activation, unlike AFB₁.97

Genotoxicity. The AFM₁ metabolite can increase the percentage of apoptotic cells,¹⁰⁶ and produce DNA damage, which leads to a prolongation of the G1 and G2 phases of the cell cycle,¹⁰⁷ in addition, AFM₁ can reduce the expression of tight junction proteins,³⁰ causing a time and concentration-

dependent reduction in cell viability,⁹⁹ significantly decreasing the S phase of the cell cycle.¹⁰⁷ Correlation between AF exposure and the appearance of guanine-aflatoxin N7 adducts in serum and urine has been demonstrated. As a consequence of DNA adduction, mutations occur during DNA repair or replication and, if the mutations are in critical genes, can significantly alter cellular functions. Studies suggest a high correlation between AFM₁ exposure and point mutations at a specific location, the third base of codon 249, of the p53 tumor suppressor gene.^{24, 108} AFM₁ does not require cytochrome P450 activation to exert cytotoxicity, therefore cells can be more easily affected by this mycotoxin in milk, and AFM₁ does not need to pass through the blood circulation and can enter milk directly.⁹⁴

Immunosuppression. AFM₁ inhibits phagocytosis and protein synthesis by interrupting the formation of DNA, RNA and proteins in the ribosome; the absorption of amino acids is altered and their hepatic retention is increased.¹⁰⁹ In innate immunity AFM₁ reduces macrophage viability, proliferation, cytotoxicity and phagocytic activity, as well as their expression of cytokines such as TNF-a, IL-1, and IL-6, and inducible nitric oxide synthase that mediate intracellular destruction of pathogens in phagocytosis.¹¹⁰ Regarding adaptive immunity, an *in vitro* study in human Jurkat lymphoblastoid T cell line, AFM₁ inhibited T cell proliferation, but did not cause apoptosis or necrosis, however, it significantly increased the expression of IL-8 involved in innate immunity, while adaptive was not affected.¹¹¹

CONCLUSION

The development of pathologies caused by ingestion or exposure to AFM_1 is a topic that is little studied, when compared to what has been reported for AFB_1 . In-depth studies are still required in the field of breast cancer and other target organs such as the kidney and lungs, as well as the intestine, although there are reports on mutagenicity and teratogenicity by exposure to AF, the exact mechanism of these pathologies closely related to AFM_1 in humans is not yet reported.

REFERENCES

- De-Rijk TC, Van-Egmond HP, Van-Der HJ, Herbes R, De-Nijs M, Samson RA, et al. A study of the 2013 western European issue of aflatoxin contamination of maize from the Balkan area. World. Mycotoxin. J. 2015;8(5):641-51.
- [2] Coffey R, Cummins E, Ward S. Exposure assessment of mycotoxins in dairy milk. Food. Control. 2009;20(3):239-49.
- [3] Govaris A, Roussi V, Koidis PA, Botsoglou NA. Distribution and stability of aflatoxin M₁ during production and storage of yogurt. Food. Addit. Contam. 2002;11(19):1043-50.
- [4] Applebaum RS, Brackett RE, Wiseman DW, Marth EH. Aflatoxin: Toxicity to dairy cattle and occurrence in milk and milk products. J Food. Prot. 1982;45(8):752-77.
- [5] Omid FM, Behdad NA, Amened O, Alí AR. The effect of different levels of aflatoxin B₁ on intestinal length, blood parameters and immune system in broiler chickens. Vet. Microbiol. 2013;9(1):73-79.
- [6] Benkerroum N. Chronic and acute toxicities of aflatoxins: Mechanisms

of action. Int. J. Environ. Res. Public. Health. 2020;17(2):423.

- [7] Sarimehmetoglu B, Kuplulu O, Haluk CT. Detection of aflatoxin M₁ in cheese samples by ELISA. Food. Control. 2004;15(1):45-9.
- [8] Ramos-Girona AJ, Marín-Sillué S, Molino-Gahete F, Vila-Donat P, Sanchis-Almenar V. Las micotoxinas: El enemigo silencioso. Arbor. 2020;196(795):540-540.
- [9] Soriano JM. Micotoxinas en alimentos. Díaz de Santos. 1st ed. España; 2015:3-58.
- [10] Nuet M. Saint Anthony tempted by lust. Two painting ways of representation in the 14th and 15th Centuries. Locus. Amoenus. 1996: 111.
- [11] Blount, WP. Turkey "X" disease. J. Br. Turkey. Fed. 1961;9:52-77.
- [12] Gimeno A. Principales factores condicionantes para el desarrollo de los hongos y la producción de micotoxinas. Ergomix. 2002.
- [13] NIH. Aflatoxinas: Sustancias cancerígenas. Instituto Nacional del Cáncer. 2018.
- [14] Rimblas-Corredor ME, Marsilla BA. Los compuestos químicos en los alimentos desde la perspectiva de la seguridad alimentaria. Histol. Histopatol. 1st ed. España; 2004: 43.
- [15] Castro E, Ahumada F. Micotoxinas: Rol e importancia en la nutrición acuícola. FOA/OMS: Fundación Chile.
- [16] Carvajal M. Transformación de la aflatoxina B₁ de alimentos, en el cancerígeno humano, aducto AFB₁-ADN. Tip. Rev. Espec. Cienc. Quim. Biol. 2013;16(2):109–20.
- [17] Martínez MM, Vargas LM, Gómez VM. Aflatoxin: Incidence, impact on health, control and prevention. Biosalud. 2013;12(2):89-109.
- [18] Sargeant K, Sheridan A, O'kelly J, Carnaghan RBA. Toxicity associated with Certain Samples of Groundnuts. Nat. 1961;192(4807): 1096-7.
- [19] Kensler TW, Roebuck BD, Wogan GN, Groopman JD. Aflatoxin: A 50-year odyssey of mechanistic and translational toxicology. Toxicol. Sci. 2011;120(1):28-48.
- [20] Van-Rensburg SJ, Cook-Mozaffari P, Van-Schalkwyk DJ, Vander Watt JJ, Vincent TJ, Purchase IF. Hepatocellular carcinoma and dietary aflatoxin in Mozambique and Transkei. Br. J. Cancer. 1985;51(5):713-26.
- [21] Nesbitt BF, O'Kelly J, Sargeant K, Sheridan A. Aspergillus flavus and turkey X disease. Toxic metabolites of Aspergillus flavus. Nature. 1962;195(4846):1062–3.
- [22] IARC. Report of the advisory group to recommend priorities for IARC Monographs on the Evaluation of carcinogenic risks to humans. Monographs during 2015-2019. IARC. 2019; 1/002.
- [23] Altamirano JR. Aflatoxinas AFM₁ en leche de consumo: Aspectos toxicológicos y metodológicos de evaluación pericial. RDU. 2019:6-16.
- [24] Haschek W, Voss K, Beasley V. Selected mycotoxins affecting animal and human health. Handb. Toxicol. Pathol. 2002;1:645-99.
- [25] Jolly PE, Inusah S, Lu B, Ellis WO, Nyarko A, Phillips TD, et al. Association between high aflatoxin B₁ levels and high viral load in VIH-positive people. World. Mycotoxin. J. 2013;6(3):255-261.
- [26] Gallo A, Moschini M, Masoero F. Aflatoxins absorption in the gastrointestinal tract and in the vaginal mucosa in lactating dairy cows. Ital. J. Anim. Sci. 2016;7(1):53-63.
- [27] Upadhaya SD, Sung HG, Lee CH, Lee SY, Kim SW, Cho KJ, et al. Comparative study on the aflatoxin B₁ degradation ability of rumen fluid from Holstein steers and Korean native goats. J. Vet. Sci. 2009;10(1) 29-34.
- [28] Moschini M, Gallo A, Piva G, Masoero F. The effects of rumen fluid on the *in vitro* aflatoxin binding capacity of different sequestering agents and in vivo release of the sequestered toxin. Anim. Feed. Sci. Technol. 2008:147(4):292-309.
- [29] Yiannikouris A, Jouany JP. Mycotoxins in feeds and their fate in animals. Anim. Res. 2002;51(2):81-99.
- [30] Wu Q, Jezkova A, Yuan Z, Pavlikova L, Dohnal V, Kuca K. Biological degradation of aflatoxins. Drug. Metab. Rev. 2009;41(1):1-7.
- [31] Min L, Fink-Gremmels J, Li D, Tong X, Tang J, Nan X, et al. An overview of aflatoxin B₁ biotransformation and aflatoxin M₁ secretion in lactating dairy cows. Anim. Nutr. 2021;7(1) 42-8.
- [32] Caruso M, Mariotti A, Zizzadoro C, Zaghini A, Ormas P, Altafini A, et al. A clonal cell line (BME-UV1) as a possible model to study bovine mammary epithelial metabolism: Metabolism and cytotoxicity of aflatoxin B₁. Toxicon. 2009;53(4):400-8.
- [33] Lindner S, Halwachs S, Wassermann L, Honscha W. Expression and subcellular localization of efflux transporter ABCG2/BCRP in

important tissue barriers of lactating dairy cows, sheep and goats. J. Vet. Pharmacol. Ther. 2013;36(6):562-70.

- [34] Nemati M, Mehran MA, Hamed PK, Masoud A. A survey on the occurrence of aflatoxin M_1 in milk samples in Ardabil, Iran. Food. Control. 2010;21(7):1022-4.
- [35] López P. Consumo diario de aflatoxinas. Gaceta UNAM. 2018.
- [36] Cravero-Ponso CF. Estabilidad de la aflatoxina M₁ (AFM₁) en leche y su reducción en el proceso de elaboración del queso ricotta. Universidad Católica Córdoba. 2017
- [37] IARC. Agents classified by the IARC monographs. Volume 1–130. Monographs on the Evaluation of carcinogenic risks to humans. IARC. 2021: 1-130.
- [38] Rangel-Muñoz EJ, Valdivia-Flores AG, Moreno-Rico O, Hernández-Delgado S, Cruz-Vázquez C, Luna-López MC de, et al. Caracterización de Aspergillus flavus y cuantificación de aflatoxinas en pienso y leche cruda de vacas en Aguascalientes. Rev. Mex. Ciencias. Pecu. 2020;11(2):435-54.
- [39] Cullen JM, Ruebner BH, Hsieh LS, Hyde DM, Hsieh DP. Carcinogenicity of Dietary Aflatoxin M₁ in male Fischer rats compared to aflatoxin B₁. Cancer. Res. 1987;47(7):1913-17.
- [40] FAO. Worldwide regulations for mycotoxins in food and feed in 2003. FAO. Italy. 2003; 81EC. Setting maximum levels for certain contaminants in foodstuffs. Official Journal of the European Union. 2006; 1881/2006.
- [41] EC. Commission Regulation (EC) No 466/2001. Setting maximum levels for certain contaminants in foodstuffs. Official Journal of the European Union. 2006; 1881/2006.
- [42] Yousof SSM, El Zubeir IEM. Chemical composition and detection of aflatoxin M₁ in camels and cow milk in Sudan. Food. Addit. Contam. Part. B, Surveill. 2020;13(4):298-304.
- [43] Udomkun P, Mutegi C, Wossen T, Atehnkeng J, Nabahungu NL, Njukwe E, et al. Occurrence of aflatoxin in agricultural produce from local markets in Burundi and Eastern Democratic Republic of Congo. Food. Sci. Nutr. 2018;6(8):2227-38.
- [44] Mohammedi-Ameur S, Dahmane M, Brera C, Kardjadj M, Ben-Mahdi MH. Occurrence and seasonal variation of aflatoxin M₁ in raw cow milk collected from different regions of Algeria. Vet. World. 2020;13(3) 433-9.
- [45] Tarannum N, Nipa MN, Das S, Parveen S. Aflatoxin M₁ detection by ELISA in raw and processed milk in Bangladesh. Toxicol. Reports. 2020;7:1339-43.
- [46] Dos Santos JS, França VR, Katto S, Santana EH. Aflatoxin M_1 in pasteurized, UHT milk and milk powder commercialized in Londrina, Brazil and estimation of exposure. Arch. Latinoam. Nutr. 2015.65(3):181-5.
- [47] Venâncio RL, Ludovico A, de Santana EH, De-Toledo EA, De-Almeida RF, Dos-Santos JS. Occurrence and seasonality of aflatoxin M₁ in milk in two different climate zones. J. Sci. Food. Agric. 2019;99(6):3203-6.
- [48] Conteçotto AC, Pante GC, Castro JC, Souza AA, Lini RS, Romoli JCZ, et al. Occurrence, exposure evaluation and risk assessment in child population for aflatoxin M₁ in dairy products in Brazil. Food. Chem. Toxicol. 2021; 148.
- [49] Foerster C, Muñoz K, Delgado-Rivera L, Rivera A, Cortés S, Müller A, et al. Occurrence of relevant mycotoxins in food commodities consumed in Chile. Mycotoxin. Res. 2020; 36(1): 63–72.
- [50] Shi R, Yu Z, Ho H, Wu W, Wang J, Tian X, et al. Survey of aflatoxin M₁ in commercial liquid milk products in China. J. Food. Prot. 2021;84(2):200-3.
- [51] Li S, Min L, Wang G, Li D, Zheng N, Wang J. Occurrence of aflatoxin M₁ in raw milk from manufacturers of infant milk powder in China. Int. J. Environ. Res. Public. Health. 2018;15(5).
- [52] Molina A, Chavarria G, Alfaro-Cascante M, Leyva A, Granados-Chinchilla F. Mycotoxins at the start of the food chain in Costa Rica: Analysis of six fusarium toxins and ochratoxin a between 2013 and 2017 in animal feed and aflatoxin M₁ in dairy products. Toxins. 2019;11(6).
- [53] Puga-Torres B, Salazar D, Cachiguango M, Cisneros G, Gómez-Bravo C. Determination of aflatoxin M₁ in raw milk from different provinces of Ecuador. Toxins. 2020;12(8):498.
- [54] Abdallah MF, Girgin G, Baydar T. Mycotoxin detection in maize, commercial feed, and raw dairy milk samples from Assiut city, Egypt. Vet. Sci. 2019;6(2).
- [55] Zakaria AM, Amin YA, Khalil OS, Abdelhiee EY, Elkamshishi MM.

Rapid detection of aflatoxin M_1 residues in market milk in Aswan Province, Egypt and effect of probiotics on its residues concentration. J. Adv. Vet. Anim. Res. 2019;6(2):197-201.

- [56] Ismaiel AA, Tharwat NA, Sayed MA, Gameh SA. Two-year survey on the seasonal incidence of aflatoxin M₁ in traditional dairy products in Egypt. J. Food. Sci. Technol. 2020;57(6):2182-9.
- [57] Mwanza M, Abdel-Hadi A, Ali AM, Egbuta M. Evaluation of analytical assays efficiency to detect aflatoxin M₁ in milk from selected areas in Egypt and South Africa. J. Dairy. Sci. 2015;98(10):6660-7.
- [58] Peña-Rodas O, Martinez-Lopez R, Hernandez-Rauda R. Occurrence of aflatoxin M₁ in cow milk in El Salvador: Results from a two-year survey. Toxicol. Reports. 2018;5:671-8.
- [59] Admasu FT, Melak A, Demissie B, Yenew C, Habtie ML, Bekele TT, et al. Occurrence and associated factors of aflatoxin M₁ in raw cow milk in South Gondar Zone, North West Ethiopia, 2020. Food. Sci. Nutr. 2021;9(11):6286-93.
- [60] Pandey AK, Shakya S, Patyal A, Ali SL, Bhonsle D, Chandrakar C, et al. Detection of aflatoxin M₁ in bovine milk from different agroclimatic zones of Chhattisgarh, India, using HPLC-FLD and assessment of human health risks. Mycotoxin. Res. 2021;37(3):265-73.
- [61] Sharma H, Jadhav VJ, Garg SR. Aflatoxin M_1 in milk in Hisar city, Haryana, India and risk assessment. Food. Addit. Contam. 2020;13(1):59-63.
- [62] Patyal A, Gill JPS, Bedi JS, Aulakh RS. Potential risk factors associated with the occurrence of aflatoxin M₁ in raw milk produced under different farm conditions. J. Environ. Sci. Health. B. 2020;55(9):827-34.
- [63] Gummadidala PM, Omebeyinje MH, Burch JA, Chakraborty P, Biswas PK, Banerjee K, et al. Complementary feeding may pose a risk of simultaneous exposures to aflatoxin M₁ and deoxynivalenol in Indian infants and toddlers: Lessons from a mini-survey of food samples obtained from Kolkata, India. Food. Chem. Toxicol. 2019;123:9-15.
- [64] Mozaffari Nejad AS, Heshmati A, Ghiasvand T. The Occurrence and risk assessment of exposure to aflatoxin M₁ in ultra-high temperature and pasteurized milk in Hamadan Province of Iran. Osong. Public. Health. Res. Perspect. 2019;10(4):22833.
- [65] Fallah AA, Barani A, Nasiri Z. Aflatoxin M₁ in raw milk in Qazvin Province, Iran: a seasonal study. Food. Addit. Contam. 2015;8(3):195-8.
- [66] Khaneghahi AH, Bahonar A, Noori N, Yazdanpanah H, Shojaee AM. Aflatoxin M₁ in raw, pasteurized and UHT milk marketed in Iran. Food. Addit. Contam. 2019;12(4):236-44.
- [67] Milićević D, Petronijević R, Petrović Z, Djinović-Stojanović J, Jovanović J, Baltić T, et al. Impact of climate change on aflatoxin M₁ contamination of raw milk with special focus on climate conditions in Serbia. J. Sci. Food. Agric. 2019;99(11):5202-10.
- [68] Roila R, Branciari R, Verdini E, Ranucci D, Valiani A, Pelliccia A, et al. A study of the occurrence of aflatoxin M₁ in milk supply chain over a seven-year period (2014-2020): Human exposure assessment and risk characterization in the population of central Italy. Foods. 2021;10(7).
- [69] Armorini S, Altafini A, Zaghini A, Roncada P. Occurrence of aflatoxin M₁ in conventional and organic milk offered for sale in Italy. Mycotoxin. Res. 2016;32(4):237-46.
- [70] Campone L, Piccinelli AL, Celano R, Pagano I, Di Sanzo R, Carabetta S, et al. Occurrence of aflatoxin M₁ in milk samples from Italy analysed by online-SPE UHPLC-MS/MS. Nat. Prod. Res. 2018;32(15):1803-8.
- [71] Serraino A, Bonilauri P, Kerekes K, Farkas Z, Giacometti F, Canever A, et al. Occurrence of aflatoxin M₁ in raw milk marketed in Italy: exposure assessment and risk characterization. Front. Microbiol. 2019; 10.
- [72] Kuboka MM, Imungi JK, Njue L, Mutua F, Grace D, Lindahl JF. Occurrence of aflatoxin M₁ in raw milk traded in peri-urban Nairobi, and the effect of boiling and fermentation. Infect Ecol. Epidemiol. 2019;9(1).
- [73] Njombwa CA, Moreira V, Williams C, Aryana K, Matumba L. Aflatoxin M₁ in raw cow milk and associated hepatocellular carcinoma risk among dairy farming households in Malawi. Mycotoxin. Res. 2020;37(1):89-96.
- [74] Hernández-Falcón TA, Monter-Arciniega A, Cruz-Cansino ND, Alanís-García E, Rodríguez-Serrano GM, Castañeda-Ovando A, et al. Effect of thermoultrasound on aflatoxin M₁ levels, physicochemical and microbiological properties of milk during storage. Ultrason. Sonochem. 2018;48.
- [75] Tahira I, Sultana N, Munir A, Hasan SM, Hanif NQ. Report:

occurrence of aflatoxin M_1 in raw and processed milk consumed in Pakistan. Pak. J. Pharm. Sci. 2019;32(3):1097-101.

- [76] Ahmad M, Awais M, Ali SW, Ali-Khan HA, Riaz M, Sultan A, et al. Occurrence of aflatoxin M₁ in raw and processed milk and assessment of daily intake in Lahore, Multan cities of Pakistan. Food. Addit. Contam. 2019;12(1):18-23.
- [77] Akbar N, Nasir M, Naeem N, Ahmad MU, Iqbal S, Rashid A, et al. Occurrence and seasonal variations of aflatoxin M_1 in milk from Punjab, Pakistan. Toxins. 2019;11(10).
- [78] Miocinovic J, Keskic T, Miloradovic Z, Kos A, Tomasevic I, Pudja P. The aflatoxin M₁ crisis in the Serbian dairy sector: the year after. Food. Addit. Contam. 2017;10(1):1-4.
- [79] Zhang J, Zheng N, Liu J, Li FD, Li SL, Wang JQ. Aflatoxin B₁ and aflatoxin M₁ induced cytotoxicity and DNA damage in differentiated and undifferentiated Caco-2 cells. Food. Chem. Toxicol. 2015;83:54-60.
- [80] Milićević D, Petronijević R, Petrović Z, Đjinović-Stojanović J, Jovanović J, Baltić T, et al. Impact of climate change on aflatoxin M₁ contamination of raw milk with special focus on climate conditions in Serbia. J. Sci. Food. Agric. 2019;99(11):5202-10.
- [81] Flores-Flores ME, González-Peñas E. Short communication: Analysis of mycotoxins in Spanish milk. J. Dairy. Sci. 2018;101(1):113–7.
- [82] Bervis N, Lorán S, Juan T, Carramiñana JJ, Herrera A, Ariño A, et al. Field monitoring of aflatoxins in feed and milk of high-yielding dairy cows under two feeding systems. Toxins. 2021;13(3).
- [83] Capelli A, Suárez G, García y Santos C, Capelli A, Suárez G, García y Santos C. Aflatoxinas en alimentos y leche de vacas de 18 establecimientos comerciales de las regiones centro-sur y este de Uruguay. Vet. 2019;55(212):52–6.
- [84] Cui X, Muhammad I, Li R, Jin H, Guo Z, Yang Y, et al. Development of a UPLC-FLD method for detection of aflatoxin B₁ and M₁ in animal tissue to study the effect of curcumin on mycotoxin clearance rates. Front. Pharmacol. 2017; 8:650.
- [85] Hayes RB, Van NJ, Raatgever JW, Ten KF. Aflatoxin exposures in the industrial setting: an epidemiological study of mortality. Food. Chem. Toxicol. 1984;22(1):39-43.
- [86] Dvorackova I, Stora C, Ayraud N. Evidence of aflatoxin B₁ in two cases of lung cancer in man. J. Cancer. Res. Clin. Oncol. 1981;100(2):221-4.
- [87] Kelly JD, Eaton DL, Guengerich FP, Coulombe RA. Aflatoxin B₁ activation in human lung. Toxicol. Appl. Pharmacol. 1997;144(1):88-95.
- [88] Stewart RJ, Pfukenyi D. Detection and levels of aflatoxin M₁ in raw milk of dairy cows from selected small scale and commercial farms in Harare, Zimbabwe Vet. J. 2016;34(1):1–6.
- [89] Marchese S, Sorice A, Ariano A, Florio S, Budillon A, Costantini S, et al. Evaluation of aflatoxin M_1 effects on the metabolomic and cytokinomic profiling of a hepatoblastoma cell line. Toxins. 2018;10(11):436.
- [90] McGlynn KA, London WT. Epidemiology and natural history of hepatocellular carcinoma. Best. Pract. Res. Clin. Gastroenterol. 2005;19(1):3-23.
- [91] Fouad AM, Ruan D, El-Senousey HK, Chen W, Jiang S, Zheng C. Harmful effects and control strategies of aflatoxin B₁ produced by *Aspergillus flavus* and *Aspergillus parasiticus* strains on poultry. Toxins. 2019;11(3):176.
- [92] Ali I, Saleem K, Wesselinova D, Haque A. Synthesis DNA binding, haemolytic and anti-cancer assays of curcumin I-based ligands and their ruthenium (III) complexes. Med. Chem. Res. 2012;22(3):1386-98.
- [93] Ali I, Lone MN, Aboul-Enein HY. Imidazoles as potential anticancer agents. RSC. Med. Chem. 2017;8(9):1742-73.
- [94] Wu K, Jia S, Zhang J, Zhang C, Wang S, Rajput SA, et al. Transcriptomics and flow cytometry reveals the cytotoxicity of aflatoxin B₁ and aflatoxin M₁ in bovine mammary epithelial cells. Ecotoxicol. Environ. Saf. 2021;209:111-823.
- [95] Creppy EE. Update of survey, regulation and toxic effects of mycotoxins in Europe. Toxicol. Lett. 2002:19-28.
- [96] Henry SH, Whitaker T, Rabbani I, Bowers J, Park D, Price W, et al. Aflatoxin M₁, JECFA. INCHEM. 2001;47(1).
- [97] Neal GE, Eaton DL, Judah DJ, Verma A. Metabolism and toxicity of aflatoxins M₁ and B₁ in human-derived in vitro systems. Toxicol. Appl. Pharmacol. 1998;151(1):152-8.
- [98] Caloni F, Stammati A, Friggè G, De Angelis I. Aflatoxin M1 absorption

and cytotoxicity on human intestinal *in vitro* model. Toxicon. 2006;47(4):409-15.

- [99] Zhang J, Zheng N, Liu J, Li FD, Li SL, Wang JQ. Aflatoxin B₁ and aflatoxin M₁ induced cytotoxicity and DNA damage in differentiated and undifferentiated Caco-2 cells. Food. Chem. Toxicol. 2015;83:54-60.
- [100] Guerra MC, Galvano F, Bonsi L, Speroni E, Costa S, Renzulli C, et al. Cyanidin-3-O-β-glucopyranoside, a natural free-radical scavenger against aflatoxin B₁- and ochratoxin A induced cell damage in a human hepatoma cell line (Hep G2) and a human colonic adenocarcinoma cell line (CaCo-2). Br. J. Nutr. 2005;94(2):211-20.
- [101] Gao Y, Bao X, Meng L, Liu H, Wang J, Zheng N. Aflatoxin B₁ and Aflatoxin M₁ induce compromised intestinal integrity through clathrinmediated endocytosis. Toxins. 2021;13(3):184.
- [102] Li H, Xing L, Zhang M, Wang J, Zheng N. The toxic effects of aflatoxin B₁ and aflatoxin M₁ on kidney through regulating L-proline and downstream apoptosis. Biomed. Res. Int. 2018;11.
- [103] Li H, Li S, Yang H, Wang Y, Wang J, Zheng N. L-proline alleviates kidney injury caused by AFB₁ and AFM₁ through regulating excessive apoptosis of kidney cells. Toxins. 2019;11(4):226.
- [104] Zheng N, Zhang H, Li S, Wang J, Liu J, Ren H, et al. Lactoferrin inhibits aflatoxin B₁ and aflatoxin M₁ induced cytotoxicity and DNA damage in Caco-2, HEK, Hep-G2, and SK-N-SH cells. Toxicon. 2018;150:77-85.
- [105] Gao Y, Ye Q, Bao X, Huang X, Wang J, Zheng N. Transcriptomic and proteomic profiling reveals the intestinal immunotoxicity induced by aflatoxin M₁ and ochratoxin A. Toxicon. 2020;180:49-61.
- [106] Liu Y, Wang W. Aflatoxin B₁ impairs mitochondrial functions, activates ROS generation, induces apoptosis and involves Nrf2 signal pathway in primary broiler hepatocytes. Anim. Sci. J. 2016;87(12):1490-500.
- [107] Hu P, Zuo Z, Li H, Wang F, Peng X, Fang J, et al. The molecular mechanism of cell cycle arrest in the bursa of fabricius in chick exposed to Aflatoxin B₁. Sci. Reports. 2018;8(1):1-9.
- [108] IARC: Some naturally occurring substances: Food items and constituents, heterocyclic aromatic amines and mycotoxin. Volume 56. Monographs on the Evaluation of cancinigenic risks to humans.
- [109] Luis J, Rojas R, Gutiérrez TR, Orantes ZM, Cruz AM, San R, et al. Contaminación por micotoxinas de la leche y derivados lácteos Contamination by micotoxins of milk and milk products. Que hacer Científico en Chiapas. 2017;12(1):90-103.
- [110] Bianco G, Russo R, Marzocco S, Velotto S, Autore G, Severino L. Modulation of macrophage activity by aflatoxins B₁ and B₂ and their metabolites aflatoxins M₁ and M₂. Toxicon. 2012;59(6) 644-50.
- [111] Luongo D, Ruso R, Balestrieri A, Marzocco S, Bèrgamo P, Severino L. *In vitro* study of AFB₁ and AFM₁ effects on human lymphoblastoid Jurkat T-cell model. J. Immunotoxicol. 2014;11(4):353-8.